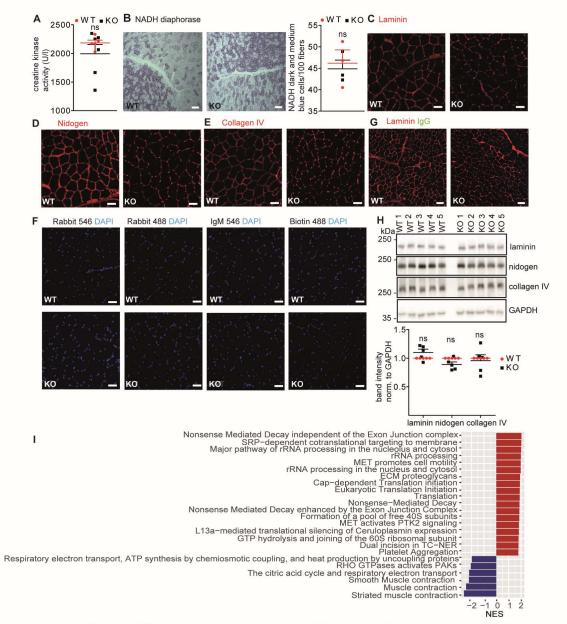


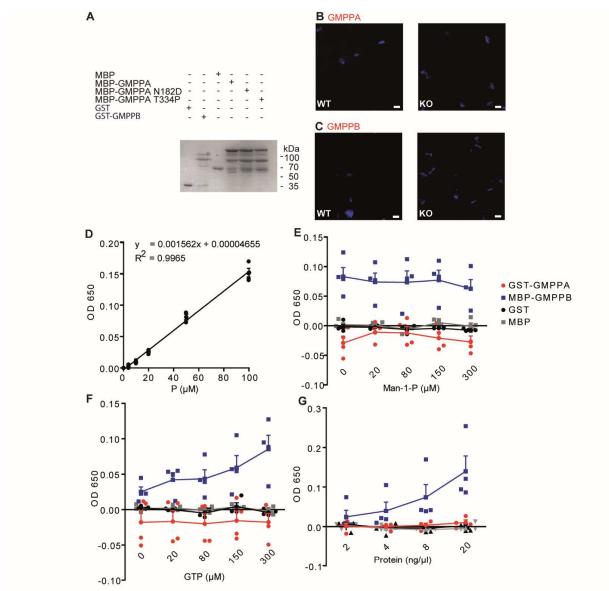
Supplementary Figure 1. Verification of *Gmppa* KO and body weight of KO mice. A) Southern blot analysis of the *Gmppa* locus of a WT and a heterozygous KO mouse exploiting the EcoRI restriction sites and the probe displayed in Figure 1A detecting a WT fragment at 6979 bp and a recombined fragment at 2671 bp. B) *Gmppa* transcript abundance determined by quantitative RT-PCR is decreased in skeletal muscle lysates of KO mice (n=3 mice per genotype; unpaired 2-tailed Student's t-test). C,D) Body weight of 3- and 12-month-old WT and KO mice for C) males and D) females (n=6 mice per genotype and age; 2-way ANOVA with Bonferoni post-hoc test). Quantitative data are presented as mean±SEM and individual data points.



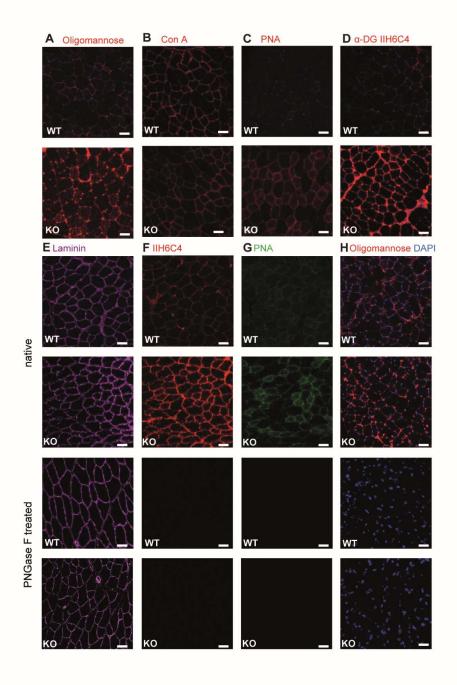
Supplementary Figure 2. Effect of *Gmppa* disruption on ECM proteins. A) The creatine kinase activity in the serum of 3-month-old KO mice is not increased (n=3 mice per group; unpaired 2-tailed Student's t-test).

B) No alteration of the oxidative potential as judged by NADH-diaphorase stainings in 12-month-old KO mice (n=4 mice per group; unpaired 2-tailed Student's t-test; scale bar: 200µm). C-E) Lower magnifications of stainings of skeletal muscle sections of 12-month-old WT and KO mice displayed in Figure 2. C) Laminin, D) Nidogen, E) Collagen IV. Scale bars C-E: 50µm. F) Control stanings of skeletal muscle sections of 12-month-old WT and KO mice stained with secondary antibodies only and DAPI to label nuclei. Scale bars: 50µm. G) The sarcolemma is intact in 12-month-old KO mice as judged by co-staining for laminin and IgG (n=3; scale bars: 100µm).

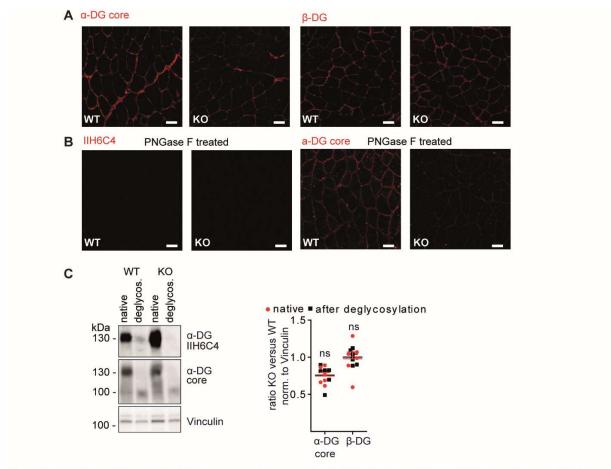
H) Nidogen and collagen IV abundances in skeletal muscle lysates are not changed at 12 months of age (n=5 mice each; 2-way ANOVA with Bonferoni post-hoc). GAPDH served as loading control (same membrane as shown in Figure 5C). I) Pathway analysis for differentially expressed proteins as determined by mass spectrometry of skeletal muscle lysates of 12-month-old WT and KO mice. Blue indicates down-regulated and red up-regulated pathways in KO mice compared to WT littermates. Data are presented as mean±SEM and individual data points.



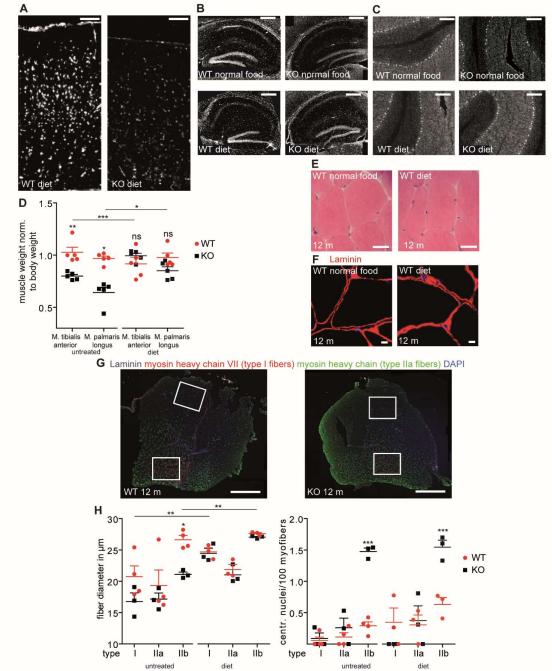
Supplementary Figure 3. GMPPA is an allosteric inhibitor of GMPPB. A) Coomassie stained gel of an aliquot of the input for the pull-down assays shown in Figure 4E. B,C) PLA assay controls. Scale bars: 5µm. B) PLA with skeletal muscle sections from Gmppa WT and KO mice with the GMPPA antibody but without the antibody directed against GMPPB. C) PLA assay with skeletal muscle sections from GMPPA WT and KO mice with the GMPPB antibody but without the antibody directed against GMPPA. D-G) Control assays for GMPPB and GMPPA activity measurements. D) The OD at 650nm is plotted as a function of the phosphate (P) concentration. E) Enzyme activities in dependence of mannose-1-phosphate concentrations. F) Enzyme activities in dependence of GTP concentrations. G) Enzymatic activities in dependence of either GMPPB or GMPPA concentrations. Quantitative data are presented as mean±SEM and individual data points from 4 independent experiments.



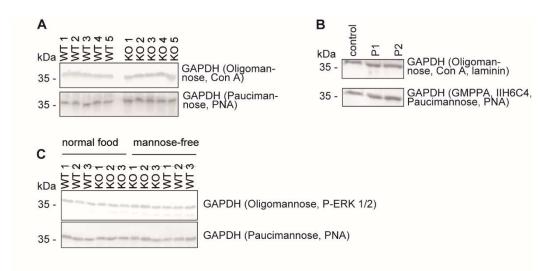
Supplementary Figure 4. Hyperglycosylation of skeletal muscle proteins in *Gmppa* KO mice. A-D) Lower magnifications of stainings of skeletal muscle sections of 12-month-old WT and KO mice displayed in Figure 5. A) Oligomannose, B) Con A, C) PNA, and D) the IIH6C4 antibody directed against the glycosylation specific epitope of α -Dg. Scale bars: 50 μ m. E-H) Stainings of skeletal muscle sections of 12-month-old WT and KO mice with the anibodies/lectins indicated. While signals for oligomannose, PNA, and the glycoslyation-specific epitope of α -DG are absent after prior PNGase F treatment, the signals for laminin are preserved. Scale bars: 50 μ m.



Supplementary Figure 5. Overviews and control stainings of skeletal muscle sections. A) Lower magnifications of stainings of skeletal muscle sections of 12-month-old WT and KO mice displayed in Figure 7. Scale bars: $50\mu m$. B) The α -DG core epitope is not masked by hyperglycosylation. Stainings of skeletal muscle sections of 12-month-old WT and KO mice. While signals for the glycoslyation-specific epitope of α -DG IIH6C4 are absent after PNGase F treatment, signals for the α -DG core protein are preserved. Scale bars: $50\mu m$. C) Western blot analysis of protein lysates of skeletal muscle specimen isolated from WT and KO mice with or without deglycosylation (n=5 mice per genotype; 2-way ANOVA with Bonferroni post-hoc test). Data are presented as mean±SEM and individual data points.



Supplementary Figure 6. Dietary intervention in WT and KO mice. A) NeuN stained section of the somatosensory cortex. Scale bars: 50μm. B) NeuN stained hippocampal section of an untreated and a treated 12-month-old WT and KO mouse. The quantification is shown in Figure 8. The image for the untreated WT is also displayed in Figure 1. Scale bars: 125μm. C) Calbindin-stained cerebellum sections of an untreated and a treated 12-month-old WT and KO mouse. The image for the untreated WT is also displayed in Figure 1. The quantification is shown in Figure 8. Scale bars: 75μm. D) Skeletal muscle weight of 12-month-old KO mice either fed on a regular or a nominally mannose-free diet (n=5 mice per group; 2-way ANOVA with Bonferoni post-hoc test). E) HE stainings of skeletal muscle sections of WT mice fed on a normal and a mannose-restricted diet. Scale bars: 50μm. The quantification is shown in Fig. 8. F) Laminin stainings of skeletal muscle sections of WT mice fed on a normal and a mannose-restricted diet. The quantification is shown in Figure 8. Scale bars: 5μm. G) Representative images of the M. gastocnemius/soleus of 12-month-old WT and KO mice. Areas used for diameter measurements are labeled. Scale bar: 1mm. H) Fiber-type specific quantifiction of fiber diameter and centralized nuclei (n=3-5 mice per group; 2-way ANOVA with Bonferoni post-hoc test). Quantitative data are presented as mean±SEM and individual data points.



Supplementary Figure 7. A) GAPDH loading controls for the membranes displayed in Figure 5. **B)** GAPDH loading controls for the membranes displayed in Figure 7. **C)** GAPDH loading controls for the membranes shown in Figure 8.